It is the season for *Cryptosporidium* and *Giardia* parasitic infections! Meridian Bioscience has provided these Technical Tips to ensure your lab gets the best possible performance when using the ImmunoCard® STAT! Crypto/Giardia test kit.

**SPECIMEN TRANSPORT**

ImmunoCard® STAT Crypto/Giardia device has been approved with the following transport media only.
- SAF
- 10% Formalin
- C&S (Cary Blair)
- MIF

Important: A 1:4 dilution of the specimen is essential. Testing specimens from overfilled vials, may cause erroneous results.

**RESULTS INTERPRETATION**

- A grey or black line in the CRYP or GIAR area, in conjunction with a line in the CONT area, should be reported as positive for *Cryptosporidium* or *Giardia* respectively.
- A line of any other color such as yellow, brown or tan should be considered invalid and the test repeated.

**KIT HANDLING & STORAGE**

To ensure optimal performance through life of the kit, ImmunoCard® STAT! Crypto/Giardia should be:
- Received cool, on an ice pack.
- Placed at 2–8°C immediately upon arrival to the facility.
- Allow kit components and specimens to equilibrate to room temperature before use.
- Returned to 2–8°C promptly after each use.

Specimens should be collected from symptomatic patients. Symptoms of *Cryptosporidium* and *Giardia* are abdominal cramps, nausea, fever and watery diarrhea.

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DILUTING UNPRESERVED SPECIMENS

Fresh specimens must be diluted 1:4 prior to testing (even if liquid-diarrheal). Dilution is necessary to obtain the same stool to diluent concentration as samples prepared in transport medium (1 part stool to 3 parts medium).

INSTRUCTIONS FOR PREPARING A 1:4 DILUTION

(mix specimens prior to diluting)

CONTAINER WITH VOLUMETRIC MARKINGS
(non-pipettable stool)
Add 3 mL of transport medium (or DI water if medium is not available) to a container with volumetric markings (test tube, beaker, graduated cylinder etc)

Add specimen until the meniscus is at 4 mL

Mix thoroughly and proceed with test

TEST TUBE
(non-pipettable stool)
Using a Precision Pipette, add 4 mL of transport medium (or DI water if medium is not available) to a test tube.

Using a permanent, marker draw a horizontal line at the miniscus of the liquid, this will be your 4 mL indicator of the test tube

Using a Precision Pipette, remove 1 mL of the DI water or transport medium from the test tube.

Add specimen to the test tube until the meniscus is at the 4 mL marking

Mix thoroughly and proceed with test

TEST TUBE
(pipettable stool)
Using a Precision Pipette, add 3 mL of transport medium (or DI water if medium is not available) to a test tube.

Using a Precision Pipette, add 1 mL of specimen

Mix thoroughly and proceed with test