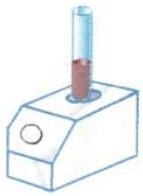
PREMIER

C. difficile GDH TEST PROCEDURE *How to perform the test*

SPECIMEN PROCESSING

1. Measure 200 µL of Sample Diluent/ Negative Control into a test tube.

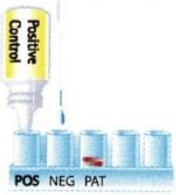
2. Mix stool thoroughly. Using a transfer pipette add 50 µL (1st mark from tip) to the tube.



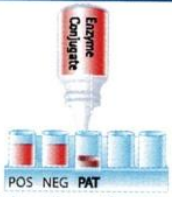
3. Vortex for 15 seconds.

SAMPLE / ENZYME CONJUGATE INCUBATION

1. Detach microwells needed, place in holder. Using transfer pipette, add 100 µL (2nd mark from tip) of the diluted stool to the appropriate wells.



2. Add 2 drops of Positive Control and 100 µL (2nd mark from tip) of Sample Diluent/Negative Control to the appropriate wells.



3. Add 1 drop of Enzyme Conjugate to all wells. Shake firmly for 30 seconds. Seal the plate and incubate at 35-39 C for 50 minutes or at 37 C for 20 minutes while rotating the plate at 1000 rpm (using the Stat-Fax rotator ONLY).

SUBSTRATE INCUBATION

1. Wash 5-7 times with 1X Wash Buffer. (See package insert for proper wash procedure.)

2. Add 2 drops of Substrate I to all wells. Shake firmly for 30 seconds and incubate for 10 minutes at 21-27 C.

3. Add 2 drops of Stop Solution I to all wells and shake firmly for 30 seconds.

INTERPRETATION OF RESULTS

Spectrophotometric Single Wavelength (450 nm)
(read within 30 minutes)
Negative = OD 450 < 0.200
Positive = OD 450 ≥ 0.200

Spectrophotometric Dual Wavelength (450/630 nm)
(read within 30 minutes)
Negative = OD 450/630 < 0.150
Positive = OD 450/630 ≥ 0.150

Please refer to the most current version of the package insert for complete instructions.

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