

PREMIER™ TOXINS A & B TEST PROCEDURE *How to perform the test*

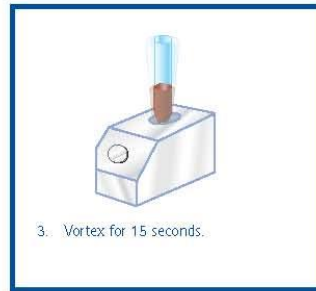
SPECIMEN PROCESSING



1. Measure 200 µL of Sample Diluent into a test tube.



2. Mix stool thoroughly. Using a transfer pipette add 50 µL (1st mark from tip) to the tube. Gently expel and withdraw the suspension several times.



3. Vortex for 15 seconds.

SAMPLE / ENZYME CONJUGATE INCUBATION



1. Detach microwells needed, place in holder. Using transfer pipette, add 100 µL (2nd mark from tip) of the diluted stool to the appropriate wells.



2. Add 2 drops of Positive Control and 100 µL (2nd mark from tip) of Negative Control (Sample Diluent) to the appropriate wells.

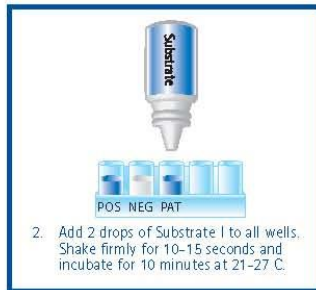


3. Add 1 drop of Enzyme Conjugate (50 µL) to all wells. Shake firmly for 30 seconds. Seal the plate and incubate at 37 °C for 50 minutes or at 37 °C for 20 minutes while rotating the plate at 1000 rpm (using the Stat-Fax rotator ONLY).

SUBSTRATE INCUBATION



1. Wash 5-7 times with 1x Wash Buffer. (See package insert for proper wash procedure.)



2. Add 2 drops of Substrate I to all wells. Shake firmly for 10-15 seconds and incubate for 10 minutes at 21-27 °C.



3. Add 2 drops of Stop Solution I to all wells and shake firmly for 30 seconds.

INTERPRETATION OF RESULTS

Visual Reading (read within 15 minutes)
 Negative = colorless to faint (barely visible) yellow
 Positive = definite yellow color

Spectrophotometric Single Wavelength (450nm)
 (read within 30 minutes)
 Negative = OD 450 < 0.150
 Positive = OD 450 ≥ 0.150

Spectrophotometric Dual Wavelength (450/630nm)
 (read within 30 minutes)
 Negative = OD 450/630 < 0.100
 Positive = OD 450/630 ≥ 0.100

* The following illustration depicts approximate sample size when testing solid, semi-solid, or non-pipetable stools.



This illustration is representative of the current Package Insert at the time of publication. Please refer to the most current version of the package insert for complete instructions.

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