Low DNA Taq HS 5 U/uL
Product Handling Guide
Shipping:

| Catalog numbers: | MDX009 |
| :--- | :--- |
| Batch No.: | See vial |
| Concentration: | $5 \mathrm{U} / \mu \mathrm{L}$ |

Store at $-20^{\circ} \mathrm{C}$

## Storage and stability:

Low DNA Taq $\mathrm{HS}, 5 \mathrm{U} / \mu \mathrm{L}$ is shipped on dry or blue ice. On arrival store at $-20^{\circ} \mathrm{C}$ for optimum stability Repeated freeze/thaw cycles should be avoided. Thawing during transportation does not affect the product performance. Solutions should be mixed/equilibrated after each thawing to avoid phasing.

## Expiry:

When stored under the recommended conditions and handled correctly, full activity of the kit is retained until the expiry date on the outer box label.

## Safety precautions:

Read and understand the SDS (Safety Data Sheets) before handling the reagents. Hardcopies of the SDSs will be provided with the first shipment, thereafter they will be available upon request.

## Quality control:

Bioline operates under ISO 13485 Management System. The Low DNA Taq HS, $5 \mathrm{U} / \mu \mathrm{L}$ and its components are extensively tested for activity, processivity, efficiency, heat activation, sensitivity, absence of nuclease contamination and absence of nucleic acid contamination.

## Notes:

This reagent has been manufactured under 13485 Quality Management System and is suitable for further manufacturing use as an IVD component.

## Description

Low DNA Taq HS, $5 \mathrm{U} / \mu \mathrm{L}$ is a highly purified and chemically modified hot-start Taq DNA polymerase. The low DNA background and stringent hot-start properties of Low DNA Taq HS, $5 \mathrm{U} / \mu \mathrm{L}$ are ideal for PCR of low-copy bacterial targets and avoiding false-positive amplification, such as in water testing.

## Kit components

Table 1

| Component |
| :--- |
| Low DNA Taq HS, $5 \mathrm{U} / \mathrm{\mu L}$ |
| Low DNA Reaction Buffer, 10x |
| 50 mM MgCl |
| 2 | Solution $\quad$.

## Users Guidelines

The optimal conditions will vary from reaction to reaction and are dependent on the template/primers used.

Pre-incubate at $95^{\circ} \mathrm{C}$ must be for 10 minutes. Subsequently, the reaction can be treated according to existing protocols.

If extension time exceeds 2.5 minutes, a maximum of 30 cycles should be used. Increasing the number of cycles may lead to smearing when run on an agarose gel.

The ideal $\mathrm{MgCl}_{2}$ concentration in the reaction is likely to be $1.5-2.5 \mathrm{mM}$ (final concentration), but some optimization may be necessary to achieve the best possible results.

## PCR reaction setup

Prepare a master mix of Low DNA Taq HS, $5 \mathrm{U} / \mu \mathrm{L}$ and assay-specific primers (see recommended composition in Table 2).

Table 2

| Reagent | Volume |
| :--- | :--- |
| Low DNA Taq HS, $5 \mathrm{U} / \mu \mathrm{L}$ | $0.4 \mu \mathrm{~L}$ |
| Low DNA Reaction Buffer, 10 x | $2 \mu \mathrm{~L}$ |
| $20 x$ Primer (\& Probe) Mix | $1.0 \mu \mathrm{~L}$ |
| 100 mM dNTP Mix | $0.2 \mu \mathrm{~L}$ |
| $50 \mathrm{mM} \mathrm{MgCl} \mathrm{I}_{2}$ Solution | $1.5 \mu \mathrm{~L}$ |
| Template | As required |
| Water $\left(\mathrm{dH}_{2} \mathrm{O}\right)$ | $\leq 20 \mu \mathrm{~L}$ |

## PCR amplification

The PCR conditions in Table 3 are suitable for amplicons of up to 1 kb .
Table 3

| Step | Temperature | Time | Cycles |
| :--- | :---: | :---: | :---: |
| Initial denaturation | $95^{\circ} \mathrm{C}$ | 10 min | 1 |
| Denaturation | $95^{\circ} \mathrm{C}$ | $0.5-1 \mathrm{~min}$ |  |
| Annealing | User <br> determined | 10 s |  |
| Extension | $72^{\circ} \mathrm{C}$ | $1 \mathrm{~min} / \mathrm{kb}$ |  |

## Technical Support

For any technical enquiries, please contact our Technical Support team via email at: mbi.tech@meridianlifescience.com

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